

SPARC and SPARC2 Flies: Inserted with CRISPR-HDR into a region near the *attP40* locus (CR-P40; see supplemental methods). **Bolded stocks** are available from the Bloomington Drosophila Stock Center (BDSC; <https://bdsc.indiana.edu/stocks/misc/sparc.html>).

1. +; ***Tl{20XUAS-SPARC-D-GCaMP6f}CR-P40***; +
2. +; ***Tl{20XUAS-SPARC-I-GCaMP6f}CR-P40***; +
3. +; ***Tl{20XUAS-SPARC-S-GCaMP6f}CR-P40***; +
4. +; ***Tl{20XUAS-SPARC-D-jGCaMP7f}CR-P40***; +
5. +; ***Tl{20XUAS-SPARC-I-jGCaMP7f}CR-P40***; +
6. +; ***Tl{20XUAS-SPARC-S-jGCaMP7f}CR-P40***; +
7. +; ***Tl{20XUAS-SPARC-I-ASAP2f}CR-P40***; +
8. +; ***Tl{20XUAS-SPARC-D-mCD8::GFP}CR-P40***; +
9. +; ***Tl{20XUAS-SPARC-I-mCD8::GFP}CR-P40***; +
10. +; ***Tl{20XUAS-SPARC-S-mCD8::GFP}CR-P40***; +
11. +; ***Tl{20XUAS-SPARC-D-LexA::p65}CR-P40***; +
12. +; ***Tl{20XUAS-SPARC-I-LexA::p65}CR-P40***; +
13. +; ***Tl{20XUAS-SPARC-S-LexA::p65}CR-P40***; +
14. +; ***Tl{20XUAS-SPARC2-D-LexA::p65}CR-P40***; +
15. +; ***Tl{20XUAS-SPARC2-I-LexA::p65}CR-P40***; +
16. +; ***Tl{20XUAS-SPARC2-S-LexA::p65}CR-P40***; +
17. +; ***Tl{20XUAS-SPARC2-D-Syn21-CsChrimson::tdTomato-3.1}CR-P40***; +
18. +; ***Tl{20XUAS-SPARC2-I-Syn21-CsChrimson::tdTomato-3.1}CR-P40***; +
19. +; ***Tl{20XUAS-SPARC2-S-Syn21-CsChrimson::tdTomato-3.1}CR-P40***; +
20. +; ***Tl{20XUAS-SPARC2-D-mCD8::GFP}CR-P40***; +
21. +; ***Tl{20XUAS-SPARC2-I-mCD8::GFP}CR-P40***; +
22. +; ***Tl{20XUAS-SPARC2-S-mCD8::GFP}CR-P40***; +

PhiC31 transgenic strains:

1. *y w; P{nSyb-IVS-PhiC31}su(Hw)attP5*; +
2. *y w; P{20XUAS-IVS-PhiC31}su(Hw)attP5*; +
3. *y w; P{αTub84-IVS-PhiC31}su(Hw)attP5*; +
4. *y w, P{nSyb-IVS-PhiC31}attP18*; +
5. *y w, P{20XUAS-IVS-PhiC31}attP18*; +
6. +, *P{nSyb-IVS-PhiC31}attP18; S/Cyo; Pr/TM6B*
7. +, *P{20XUAS-IVS-PhiC31}attP18; S/Cyo; Pr/TM6B*

Supplementary Table 5: Transgenic flies generated in this study.

R2 neuron transgene expression	Resting membrane potential (mean \pm SEM, mV)*	Input resistance (mean \pm SEM, M Ω)*
CsChrimson::tdTomato+, GFP+	-38 \pm 5	220 \pm 14
GFP+	-48 \pm 2	628 \pm 131

Supplementary Table 7: Resting membrane potential and input resistance of R2 neurons from Figure 6.

Supplementary Note - Fly genotypes (by figure):

Figure 2b: +; $Tl\{20XUAS-SPARC-D-GCaMP6f\}CR-P40/+; P\{VT025965-Gal4\}attP2/P\{10XUAS-IVS-myr::tdTomato\}attP2$

Figure 2c: +; $Tl\{20XUAS-SPARC-D-GCaMP6f\}CR-P40/P\{nSyb-IVS-PhiC31\}su(Hw)attP5; P\{VT025965-Gal4\}attP2/P\{10XUAS-IVS-myr::tdTomato\}attP2$

Figure 2d: +; $Tl\{20XUAS-SPARC-I-GCaMP6f\}CR-P40/P\{nSyb-IVS-PhiC31\}su(Hw)attP5; P\{VT025965-Gal4\}attP2/P\{10XUAS-IVS-myr::tdTomato\}attP2$

Figure 2e: +; $Tl\{20XUAS-SPARC-S-GCaMP6f\}CR-P40/P\{nSyb-IVS-PhiC31\}su(Hw)attP5; P\{VT025965-Gal4\}attP2/P\{10XUAS-IVS-myr::tdTomato\}attP2$

Figure 2g-g'': +; $Tl\{20XUAS-SPARC-I-LexA::p65\}CR-P40/+; P\{GMR19F01-Gal4\}attP2/P\{UAS-mCD8::GFP\}attP1 P\{13XlexAop-IVS-myr::tdTomato\}attP2$

Figure 2h-h'': +; $Tl\{20XUAS-SPARC2-I-LexA::p65\}CR-P40/+; P\{GMR19F01-Gal4\}attP2/P\{10XUAS-mCD8::GFP\}attP1 P\{13XlexAop-IVS-myr::tdTomato\}attP2$

Figure 2i-i'': +; $Tl\{20XUAS-SPARC2-D-LexA::p65\}CR-P40/P\{nSyb-IVS-PhiC31\}su(Hw)attP5; P\{GMR19F01-Gal4\}attP2/P\{10XUAS-IVS-mCD8::GFP\}attP1, P\{13XlexAop-IVS-myr::tdTomato\}attP2$

Figure 2j-j'': +; $Tl\{20XUAS-SPARC2-I-LexA::p65\}CR-P40/P\{nSyb-IVS-PhiC31\}su(Hw)attP5; P\{GMR19F01-Gal4\}attP2/P\{10XUAS-IVS-mCD8::GFP\}su(Hw)attP1, P\{13XlexAop-IVS-myR::tdTomato\}attP2$

Figure 2k-k'': +; $Tl\{20XUAS-SPARC2-S-LexA::p65\}CR-P40/P\{nSyb-IVS-PhiC31\}su(Hw)attP5; P\{GMR19F01-Gal4\}attP2/P\{10XUAS-IVS-mCD8::GFP\}su(Hw)attP1, P\{13XlexAop-IVS-myR::tdTomato\}attP2$

Figure 3a: +; $Tl\{20XUAS-SPARC2-D-mCD8::GFP\}CR-P40/P\{nSyb-IVS-PhiC31\}su(Hw)attP5; P\{VT025965-Gal4\}attP2/P\{10XUAS-IVS-myR::tdTomato\}attP2$

Figure 3b: +; $Tl\{20XUAS-SPARC2-I-mCD8::GFP\}CR-P40/P\{nSyb-IVS-PhiC31\}su(Hw)attP5; P\{VT025965-Gal4\}attP2/P\{10XUAS-IVS-myR::tdTomato\}attP2$

Figure 3c: +; $Tl\{20XUAS-SPARC2-S-mCD8::GFP\}CR-P40/P\{nSyb-IVS-PhiC31\}su(Hw)attP5; P\{VT025965-Gal4\}attP2/P\{10XUAS-IVS-myR::tdTomato\}attP2$

Figure 3d: +; $Tl\{20XUAS-SPARC2-D-mCD8::GFP\}CR-P40/P\{nSyb-IVS-PhiC31\}su(Hw)attP5; P\{GMR19F01-Gal4\}attP2/P\{10XUAS-IVS-myR::tdTomato\}attP2$

Figure 3e: +; $Tl\{20XUAS-SPARC2-I-mCD8::GFP\}CR-P40/P\{nSyb-IVS-PhiC31\}su(Hw)attP5; P\{GMR19F01-Gal4\}attP2/P\{10XUAS-IVS-myR::tdTomato\}attP2$

Figure 3f: +; $Tl\{20XUAS-SPARC2-S-mCD8::GFP\}CR-P40/P\{nSyb-IVS-PhiC31\}su(Hw)attP5; P\{GMR19F01-Gal4\}attP2/P\{10XUAS-IVS-myR::tdTomato\}attP2$

Figure 3g: +; $Tl\{20XUAS-SPARC2-D-mCD8::GFP\}CR-P40/P\{nSyb-IVS-PhiC31\}su(Hw)attP5; P\{GH146-Gal4\}attP2/P\{10XUAS-IVS-myR::tdTomato\}attP2$

Figure 3h: +; $Tl\{20XUAS-SPARC2-I-mCD8::GFP\}CR-P40/P\{nSyb-IVS-PhiC31\}su(Hw)attP5; P\{GH146-Gal4\}attP2/P\{10XUAS-IVS-myR::tdTomato\}attP2$

Figure 3i: +; $Tl\{20XUAS-SPARC2-S-mCD8::GFP\}CR-P40/P\{nSyb-IVS-PhiC31\}su(Hw)attP5; P\{GH146-Gal4\}attP2/P\{10XUAS-IVS-myR::tdTomato\}attP2$

Figure 3j: $P\{nSyb-IVS-PhiC31\}attP18; PBac\{R17A04-p65.ADZp\}VK00027/Tl\{20XUAS-SPARC2-D-mCD8::GFP\}CR-P40; P\{R35B06-ZpGAL4.DB\}attP2/P\{10XUAS-IVS-myR::tdTomato\}attP2$

Figure 3k: $P\{nSyb-IVS-PhiC31\}attP18; PBac\{R17A04-p65.ADZp\}VK00027/Tl\{20XUAS-SPARC2-I-mCD8::GFP\}CR-P40; P\{R35B06-ZpGAL4.DB\}attP2 / P\{10XUAS-IVS-myR::tdTomato\}attP2$

Figure 3l: $P\{nSyb-IVS-PhiC31\}attP18; PBac\{R17A04-p65.ADZp\}VK00027/Tl\{20XUAS-SPARCS-I-mCD8::GFP\}CR-P40; P\{R35B06-ZpGAL4.DB\}attP2/P\{10XUAS-IVS-myR::tdTomato\}attP2$

Figure 3m: +; $Tl\{20XUAS-SPARC2-D-mCD8::GFP\}CR-P40$ $P\{nSyb-IVS-PhiC31\}su(Hw)attP5$; $P\{GMR27B03-Gal4\}attP2/P\{10XUAS-IVS-myrl::tdTomato\}attP2$

Figure 3n: +; $Tl\{20XUAS-SPARC2-I-mCD8::GFP\}CR-P40/P\{nSyb-IVS-PhiC31\}su(Hw)attP5$; $P\{GMR27B03-Gal4\}attP2/P\{10XUAS-IVS-myrl::tdTomato\}attP2$

Figure 3o: +; $Tl\{20XUAS-SPARC2-S-mCD8::GFP\}CR-P40/P\{nSyb-IVS-PhiC31\}su(Hw)attP5$; $P\{GMR27B03-Gal4\}attP2/P\{10XUAS-IVS-myrl::tdTomato\}attP2$

Figure 4a-c': +; $Tl\{20XUAS-SPARC2-I-mCD8::GFP\}CR-P40/P\{nSyb-IVS-PhiC31\}su(Hw)attP5$; $P\{GH146-Gal4\}/P\{10XUAS-IVS-myrl::tdTomato\}attP2$

Figure 4d-g': +; $Tl\{20XUAS-SPARC2-S-mCD8::GFP\}CR-P40/P\{nSyb-IVS-PhiC31\}su(Hw)attP5$; $P\{GH146-Gal4\}/P\{10XUAS-IVS-myrl::tdTomato\}attP2$

Figure 5a-e:

SPARC: +; $Tl\{20XUAS-SPARC-S-GCaMP6f\}CR-P40/P\{nSyb-IVS-PhiC31\}su(Hw)attP5$; $P\{VT025965-Gal4\}attP2/+$

FlpOut: +/y w, $P\{hsFLP\}$; $P\{20XUAS-IVS-GCaMP6f\}attP40/P\{\alpha Tub84b\{FRT.Gal80\}\}2$; $P\{VT025965-Gal4\}attP2/+$

Figure 6b,c: $P\{nSyb\text{-IVS}\text{-}PhiC31\}attP18/+; TI\{20XUAS\text{-}SPARC2\text{-}D\text{-}Syn21\text{-}CsChrimson\text{:}\text{:}tdTomato\text{-}3.1\}CR\text{-}P40/\{20XUAS\text{-}IVS\text{-}mCD8\text{:}\text{:}GFP\}attP40; P\{GMR19C08\text{-}Gal4\}attP2/+$

Figure 6d,e: $P\{nSyb\text{-IVS}\text{-}PhiC31\}attP18/y w; TI\{3XP3\text{-}DsRed=20XUAS\text{-}SPARC2\text{-}D\text{-}Syn21\text{-}CsChrimson\text{:}\text{:}tdTomato\text{-}3.1\}CR\text{-}P40/ P\{20XUAS\text{-}IVS\text{-}mCD8\text{:}\text{:}GFP\}attP40; P\{GMR19C08\text{-}Gal4\}attP2/+$

Extended Data Figure 1c-d": +; $TI\{20XUAS\text{-}34bp_attP\text{-}inversion\text{-}GCaMP6f\}CR\text{-}P40/P\{nSyb\text{-IVS}\text{-}PhiC31\}su(Hw)attP5; P\{GMR19F01\text{-}Gal4\}attP2/P\{10XUAS\text{-}IVS\text{-}myr\text{:}\text{:}tdTomato\}attP2$

Extended Data Figure 1f-f": +; $TI\{20XUAS\text{-}Lex\text{-}OR\text{-}Flp\}CR\text{-}P40/P\{nSyb\text{-IVS}\text{-}PhiC31\}su(Hw)attP5; P\{GMR19F01\text{-}Gal4\}attP2/P\{UAS\text{-}FRT\text{-}stop\text{-}FRT\text{-}mCD8\text{:}\text{:}GFP\}, P\{13XlexAop\text{-}IVS\text{-}myr\text{:}\text{:}tdTomato\}attP2$

Extended Data Figure 3a, e-e": +/w; $TI\{20XUAS\text{-}SPARC\text{-}D\text{-}GCaMP6f\}CR\text{-}P40/P\{R19B03\text{-}p65.ADZp\}attP40; P\{R26E07\text{-}ZpGAL4.DBD\}attP2/P\{10X UAS\text{-}myr\text{:}\text{:}tdTomato\}attP2$

Extended Data Figure 3b, f-f": +; $TI\{20XUAS\text{-}SPARC\text{-}D\text{-}GCaMP6f\}CR\text{-}P40/P\{nSyb\text{-}PhiC31\}su(Hw)attP5 P\{R19B03\text{-}p65.ADZp\}attP40; P\{R26E07\text{-}ZpGAL4.DBD\}attP2/P\{10XUAS\text{-}IVS\text{-}myr\text{:}\text{:}tdTomato\}attP2$

Extended Data Figure 3c, g-g": +; $Tl\{20XUAS-SPARC-I-SPARC-GCaMP6f\}CR-P40/P\{nSyb-PhiC31\}su(Hw)attP5 P\{R19B03-p65.ADZp\}attP40; P\{R26E07-ZpGAL4.DBD\}attP2/ P\{10XUAS-IVS-myrltdTomato\}attP2$

Extended Data Figure 3d, h-h": +; $Tl\{20XUAS-SPARC-S-GCaMP6f\}CR-P40/P\{nSyb-PhiC31\}su(Hw)attP5, P\{R19B03-p65.ADZp\}attP40; P\{R26E07-ZpGAL4.DBD\}attP2/P\{10XUAS-IVS-myrltdTomato\}attP2$

Supplementary Note - Origin of transgenes

1. $P\{GMR19F01-Gal4\}attP2$ from BDSC, stock number 48852¹.
2. $P\{VT025965-Gal4\}attP2$ from VDRC².
3. $P\{GH146-Gal4\}$ ³ and $P\{UAS-FRT-stop-FRT-mCD8::GFP\}$ ⁴ were generous gifts from ChiChi Xie and Liqun Luo.
4. $P\{R19B03-p65.ADZp\}attP40; P\{R026E07-ZpGAL4.DBD\}attP2$ and $PBac\{R17A04-p65.ADZp\}VK00027; P\{R35B06-ZpGAL4.DBD\}attP2$ from Janelia FlyLight collection.
5. $P\{GMR27B03-Gal4\}attP2$ ⁵ from BDSC, stock number 49211.
6. $P\{GMR19C08-Gal4\}attP2$ from BDSC, stock number 48845, and is described in⁶.
7. $P\{10XUAS-myrltdTomato\}attP2$ from BDSC, stock number 32221.
8. $P\{13XlexAop-myrltdTomato\}attP2$ was a generous gift from Heather Dionne and Gerald Rubin.
9. $P\{10XUAS-IVS-mCD8::GFP\}su(Hw)attP1$ from BDSC, stock number 32187.

10. $P\{20XUAS-IVS-mCD8::GFP\}attP40$ was a gift from Barret Pfeiffer and Gerald Rubin and is described in⁷.
11. $CyO, P\{w[+mC]=Crew\}DH1$ from BDSC, stock number 1092, and is described in⁸.
12. $P\{20XUAS-IVS-GCaMP6f\}attP40$ from BDSC, stock number 42747, and is described in⁹.
13. $P\{hsFLP\}1$ from BDSC stock number 6, and is described in¹⁰.
14. $P\{\alpha Tub84b(FRT.Gal80)\}2$ from BDSC, stock number 38880, and is described in¹¹.

References:

1. Jenett, A. et al. A GAL4-driver line resource for Drosophila neurobiology. *Cell Reports* **2**, 991–1001 (2012).
2. Leonhardt, A. et al. Asymmetry of Drosophila ON and OFF motion detectors enhances real-world velocity estimation. *Nat Neurosci* **19**, 706–715 (2016).
3. Nern, A., Pfeiffer, B. D. & Rubin, G. M. Optimized tools for multicolor stochastic labeling reveal diverse stereotyped cell arrangements in the fly visual system. *Proc. Natl. Acad. Sci. U.S.A.* **112**, E2967–76 (2015).
4. Hong, W. et al. Leucine-rich repeat transmembrane proteins instruct discrete dendrite targeting in an olfactory map. *Nat Neurosci* **12**, 1542–1550 (2009).
5. Dionne, H., Hibbard, K. L., Cavallaro, A., Kao, J.-C. & Rubin, G. M. Genetic Reagents for Making Split-GAL4 Lines in Drosophila. *Genetics* **209**, 31–35 (2018).
6. Jenett, A. et al. A GAL4-Driver Line Resource for Drosophila Neurobiology. *Cell Reports* **2**, 991–1001 (2012).
7. Pfeiffer, B. D. et al. Refinement of Tools for Targeted Gene Expression in Drosophila. *Genetics* **186**, 735–755 (2010).
8. Thorpe, H. M. & Smith, M. C. In vitro site-specific integration of bacteriophage DNA catalyzed by a recombinase of the resolvase/invertase family. *PNAS* **95**, 5505–5510 (1998).
9. Chen, T.-W. et al. Ultrasensitive fluorescent proteins for imaging neuronal activity. *Nature* **499**, 295–300 (2013).
10. Xu, T. & Rubin, G. M. Analysis of genetic mosaics in developing and adult Drosophila tissues. *Development* **117**, 1223–1237 (1993).

11. Del-Valle-Rodriguez, A., Didiano, D. & Desplan, C. Power tools for gene expression and clonal analysis in *Drosophila*. *Nat. Methods* **9**, 47–55 (2011).